

# Atypical periodic paralysis and myalgia

## A novel *RYR1* phenotype

Emma Matthews, MRCP, Christoph Neuwirth, MD, Fatima Jaffer, MRCP, Renata S. Scalco, MD, Doreen Fialho, MRCP, Matt Parton, FRCP, Dipa Raja Rayan, MRCP, Karen Suetterlin, MRCP, Richa Sud, PhD, Roland Spiegel, MD, Rachel Mein, BSc, Henry Houlden, FRCP, Andrew Schaefer, MRCP, Estelle Healy, FRCPPath, Jacqueline Palace, FRCP, Ros Quinlivan, FRCP, Susan Treves, PhD, Janice L. Holton, FRCPPath, Heinz Jungbluth, PhD,\* and Michael G. Hanna, FRCP\*

### Correspondence

Dr. Matthews  
emma.matthews@ucl.ac.uk

*Neurology*® 2018;90:e1-7. doi:10.1212/WNL.0000000000004894

## Abstract

### Objective

To characterize the phenotype of patients with symptoms of periodic paralysis (PP) and ryanodine receptor (*RYR1*) gene mutations.

### Methods

Cases with a possible diagnosis of PP but additional clinicopathologic findings previously associated with *RYR1*-related disorders were referred for a tertiary neuromuscular clinical assessment in which they underwent detailed clinical evaluation, including neurophysiologic assessment, muscle biopsy, and muscle MRI. Genetic analysis with next-generation sequencing and/or targeted Sanger sequencing was performed.

### Results

Three cases with episodic muscle paralysis or weakness and additional findings compatible with a *RYR1*-related myopathy were identified. The McManis test, used in the diagnosis of PP, was positive in 2 of 3 cases. Genetic analysis of known PP genes was negative. *RYR1* analysis confirmed likely pathogenic variants in all 3 cases.

### Conclusions

*RYR1* mutations can cause late-onset atypical PP both with and without associated myopathy. Myalgia and cramps are prominent features. The McManis test may be a useful diagnostic tool to indicate *RYR1*-associated PP. We propose that clinicopathologic features suggestive of *RYR1*-related disorders should be sought in genetically undefined PP cases and that *RYR1* gene testing be considered in those in whom mutations in *SCN4A*, *CACNA1S*, and *KCNJ2* have already been excluded.

\*These authors contributed equally to this work.

From the MRC Centre for Neuromuscular Diseases (E.M., F.J., R.S.S., D.F., M.P., D.R.R., K.S., H.H., E.H., R.Q., J.L.H., M.G.H.), Department of Molecular Neuroscience, UCL Institute of Neurology and National Hospital for Neurology and Neurosurgery, Queen Square, London, UK; Neuromuscular Diseases Unit/ALS Clinic (C.N.), Kantonsspital St. Gallen, Switzerland; Neurogenetics Unit (R.S., H.H.) and Department of Neuropathology (J.L.H.), National Hospital for Neurology and Neurosurgery, Queen Square, London, UK; Human Genetics Laboratory Genetica (R.S.), Zurich, Switzerland; Genetics Department (R.M.), Viapath, Guy's Hospital, London; Wellcome Trust Centre for Mitochondrial Research (A.S.), University of Newcastle, Framlington Place, Newcastle Upon Tyne, UK; Institute of Pathology (E.H.), Belfast Health and Social Care Trust, Northern Ireland; Department of Neurology (J.P.), John Radcliffe Hospital, Oxford, UK; Departments of Biomedicine and Anesthesia (S.T.), Basel University Hospital, Switzerland; Department of Life Sciences (S.T.), Microbiology and Applied Pathology Section, University of Ferrara, Italy; Department of Paediatric Neurology (H.J.), Neuromuscular Service, Evelina Children's Hospital, St. Thomas' Hospital; and Department of Basic and Clinical Neuroscience (H.J.), Institute of Psychiatry, Psychology and Neuroscience, and Randall Division of Cell and Molecular Biophysics (H.J.), Muscle Signalling Section, King's College, London, UK.

Go to [Neurology.org/N](http://Neurology.org/N) for full disclosures. Funding information and disclosures deemed relevant by the authors, if any, are provided at the end of the article.

The Article Processing Charge was funded by the Medical Research Council (UK).

This is an open access article distributed under the terms of the Creative Commons Attribution License 4.0 (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## Glossary

**CAMP** = compound muscle action potential; **MH** = malignant hyperthermia; **MmD** = multiminicore disease; **PEG** = percutaneous endoscopic gastrostomy; **PP** = periodic paralysis; **RYR1** = ryanodine receptor.

The skeletal muscle ryanodine receptor (*RYR1*) gene encodes the principal sarcoplasmic reticulum calcium release channel with a crucial role in excitation-contraction coupling. Mutations in *RYR1* are the most common genetic cause of nondystrophic neuromuscular disorders,<sup>1,2</sup> associated with a wide spectrum of clinicopathologic features, ranging from various early-onset congenital myopathies—central core disease,<sup>3</sup> multiminicore disease (MmD),<sup>4</sup> centronuclear myopathy,<sup>5</sup> and congenital fiber type disproportion<sup>6</sup>—to the malignant hyperthermia (MH) susceptibility trait. *RYR1* mutations may also give rise to episodic neuromuscular manifestations, including exertional myalgia and rhabdomyolysis,<sup>7</sup> a late-onset axial myopathy,<sup>8,9</sup> and have recently been associated with a novel bleeding disorder due to abnormal smooth muscle cell contractility.<sup>10</sup> Episodes of atypical periodic paralysis (PP) have been previously reported in a single patient with a recessive *RYR1*-related myopathy,<sup>11</sup> but it has been unclear whether this phenotype was unique to the reported individual or common across different *RYR1* genotypes. The association between *RYR1* and PP is not entirely unexpected, considering that dysfunction of Cav1.1, the other key player involved in excitation-contraction coupling and the principal *RYR1* interactor, is the most common cause of primary PP. Recently, both dominant and recessive families with a Cav1.1-related myopathy have been described,<sup>12</sup> suggesting further phenotypic overlap between *RYR1* and Cav1.1 dysfunction and a continuum between myopathic and episodic phenotypes due to mutations in these genes. Here, we report 3 additional *RYR1*-mutated patients presenting with PP episodes and variable additional myopathic manifestations.

## Methods

We examined cases referred with a possible diagnosis of PP but additional clinicopathologic findings previously associated with *RYR1*-related disorders to the national referral center for skeletal muscle channelopathies in the United Kingdom and to the Swiss Neuromuscular Diseases Unit Center.

### Standard protocol approvals, registrations, and patient consents

All procedures were conducted as part of routine clinical care. The study was performed under the ethics guidelines issued by our institution, with written informed consent obtained from all participants for genetic studies.

Genetic analysis for PP genes *SCN4A*, *CACNA1S*, and *KCNJ2* was performed at the Neurogenetics Unit, National Hospital for Neurology and Neurosurgery as provided by the

Channelopathy Highly Specialized National Service for rare disease. Samples underwent next-generation sequencing on an Illumina HiSeq after enrichment with an Illumina custom Nextera Rapid Capture panel (Illumina, Inc, San Diego, CA). For case 2, the library preparation and enrichment were performed with TruSight One kit (Illumina) according to protocol instructions, allowing analysis of a panel of ≈5,000 genes (including PP genes and *RYR1*). The library was quantified with the Qubit 2.0 Fluorometer system (Thermo Fisher Scientific, Waltham, MA), and 2 × 250-bp paired-end sequencing was performed on the MiSeq sequencer (Illumina), as well as sequences alignment (Burrows-Wheeler aligner) and variant calling (Genome Analysis Toolkit variant caller). The variants were then analyzed with VariantStudio (Illumina).

Additional targeted *RYR1* Sanger sequencing of all cases was performed at the Diagnostic DNA Laboratory at Guy's Hospital, London.

## Results

### Case 1

A 54-year-old man was born 7 weeks prematurely. An atrial septal defect was noted at birth and corrected at the age of 5 years. Walking was delayed until the age of 2 years. He was never able to run or to keep up with peers physically because of weakness of his arms and legs. In early childhood, he toe-walked, ultimately requiring Achilles tendon lengthening. Symptoms were effectively stable throughout childhood, but from the age of 20 years, there was slowly progressive proximal weakness.

At the age of 34 years, after a flu-like illness, he complained of a change in symptoms. He reported episodes of sudden severe myalgia followed by profound muscle weakness in either 1 limb or the entire body from the neck down lasting for several hours. Examination demonstrated a waddling gait and pronounced lumbar lordosis and mild dysmorphic features with a long thin face and high arched palate. Mild bilateral ptosis with a complex ophthalmoplegia most marked on upgaze, a typical finding in recessive *RYR1*-related myopathies, was noted. There was bilateral facial and sternocleidomastoid weakness with additional proximal upper and lower limb weakness. Reflexes were 1+, and there were no sensory abnormalities. Cardiac examination demonstrated atrial fibrillation for which he underwent cardioversion. Because of the change in symptoms, a muscle biopsy was performed. Although core-like structures were seen, they were not felt to be typical of central core disease, a common

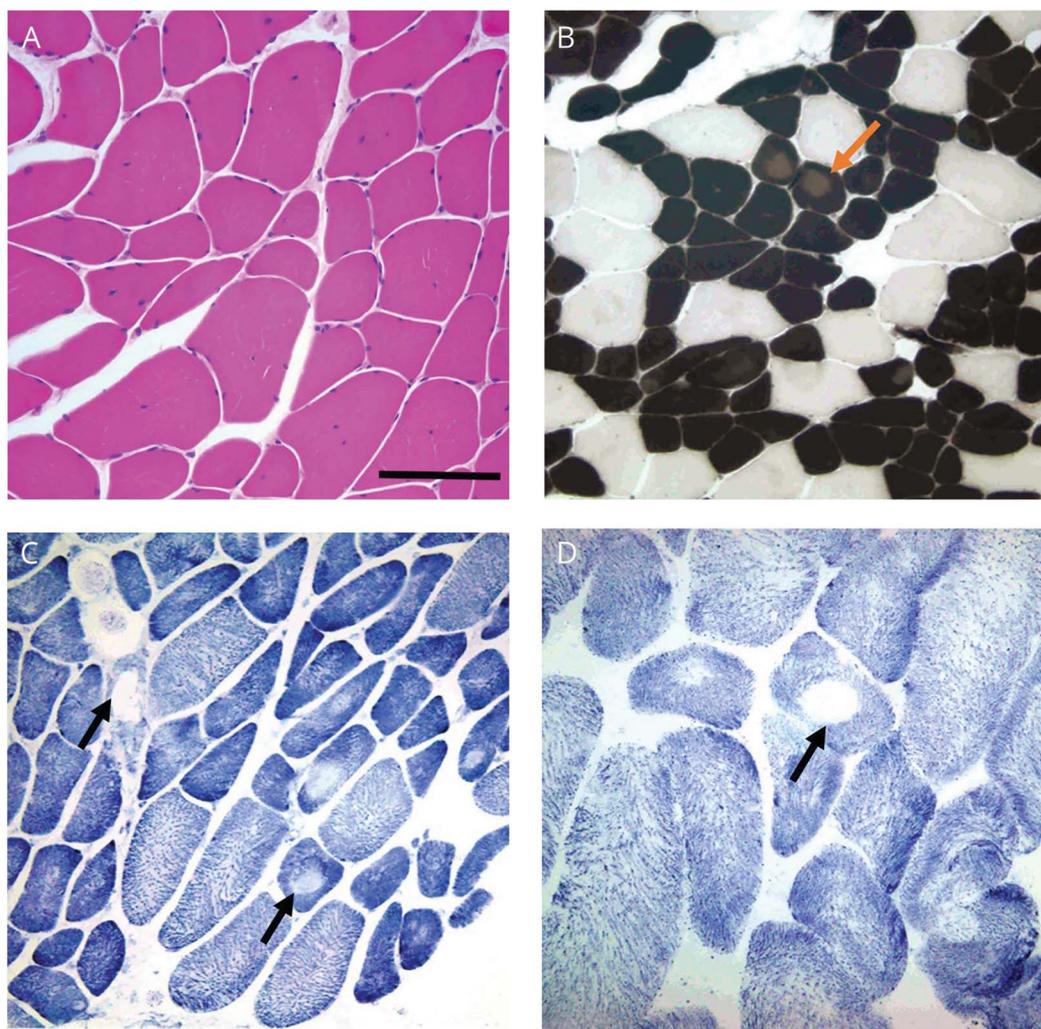
*RYR1*-related myopathy, and no definitive diagnosis was reached (figure 1).

From this point, he continued to complain of episodes of temporary worsening of limb weakness 2 to 3 times a year lasting for several days. He could identify no specific triggers. At the age of 38 years, he presented with an episode of severe neck flexor and bulbar weakness. Over the course of 9 days, his neck flexor weakness improved to baseline. Bulbar function remained impaired, and a percutaneous endoscopic gastrostomy (PEG) tube for feeding was inserted. However, he continued to report fluctuant improvement in his swallow, on some days being entirely reliant on the PEG and on others managing a soft diet. Fourteen months after the PEG insertion, he attended clinic and reported that he no longer used it and consistently managed a normal diet. Videofluoroscopy demonstrated only minor persisting abnormalities, and the PEG was removed at his request.

Over subsequent years, he continued to experience acute attacks of myalgia followed by weakness. When care was transferred to us, he was reinvestigated, including a repeat muscle biopsy at the age of 42 years (figure 2), which on this occasion revealed mild loss of oxidative enzyme activity in the central region of many fibers, but classic central cores were not observed.

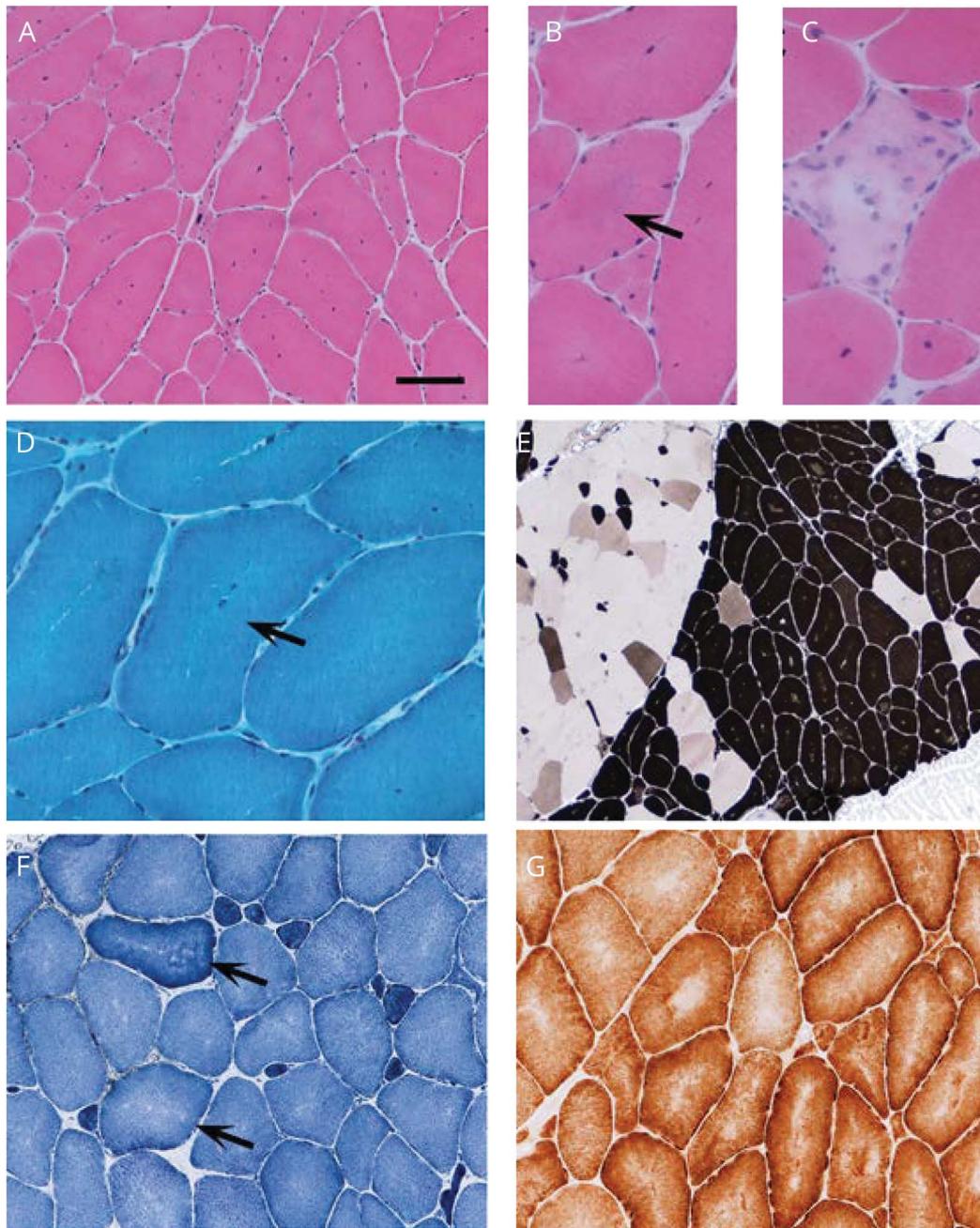
Electrophysiologic tests showed myopathic EMG features, normal repetitive stimulation, mildly abnormal jitter, and occasional blocking on single-fiber EMG (including in extensor digitorum communis, biceps, and orbicularis oculi). While indicative of a mild neuromuscular junction transmission defect, this was thought likely to be a secondary phenomenon rather than a primary myasthenic feature.<sup>13</sup> Serologic testing for acetylcholine receptor antibodies and muscle-specific kinase antibodies was negative. The McManis test for PP was positive on 2 separate occasions (53% and 59% decrement in

**Figure 1** Histology slides of muscle biopsies taken from case 1 at age (A) 34 and (B) 42 years



(A) Histologic examination of a right quadriceps muscle biopsy performed at age 34 years showed variation in fiber diameter and internalized nuclei with hematoxylin and eosin staining. (B) ATPase histochemistry at pH 4.3 indicated type I fiber predominance with decreased myofibrillar ATPase activity in core-like areas in up to 15% of type I fibers (red arrow). (C) Central absence of staining suggestive of cores was apparent in type I and II fibers in the nicotinamide dinucleotide tetrazolium reductase preparation (black arrows), and central reduction in oxidative enzyme activity was confirmed by succinate dehydrogenase staining (D, black arrow). Scale bar in A represents 100  $\mu$ m in (A) and (C), 200  $\mu$ m in (B), and 50  $\mu$ m in (D).

**Figure 2** Histologic examination of a right biceps muscle biopsy



Histologic examination of a right biceps muscle biopsy performed at age 42 years showed (A) variation in fiber diameter and increased internal nuclei with (B) occasional fibers showing central basophilia (arrow) and (C) a small number of necrotic fibers. Gomori trichrome staining suggested regions with reduced mitochondrial staining (arrow). (E) ATPase histochemistry indicated focal type I fiber predominance (darkly stained fibers). (F) Mild central pallor suggestive of cores was apparent in type I and II fibers in the nicotinamide adenine dinucleotide tetrazolium reductase preparation (arrows), and (G) central reduction in oxidative enzyme activity was confirmed by cytochrome oxidase histochemistry. Scale bar in represents 260  $\mu\text{m}$  in (E); 100  $\mu\text{m}$  in (A), (F), and (G); and 50  $\mu\text{m}$  in (B–D).

compound muscle action potential [CMAP] amplitude). MRI demonstrated fatty infiltration of all thigh muscles, in particular the adductor magnus, which was almost completely replaced by fat, with relative sparing of the vastus lateralis, rectus femoris, gracilis, and semitendinosus, a pattern of selective involvement previously associated with *RYR1*-related myopathies.<sup>14,15</sup> There was no family history of similar symptoms. Genetic analysis revealed *RYR1* variants Arg109Trp, previously described with MmD and ophthalmoplegia,<sup>4,16</sup> a variant of uncertain

significance Met485Val,<sup>4</sup> and the novel although presumed truncating variant Gln70X. Testing for congenital myasthenia and PP genes was negative. His unaffected mother carried the Gln70X variant. It was not possible to obtain DNA from his father.

### Case 2

A 42-year-old Swiss woman presented with intermittent weakness of the limbs lasting from several minutes to 2 days.

Medical history was notable for migraine and 3 caesarean sections. The first episode of flaccid paresis occurred during her first pregnancy at the age of 23 years with a fall from the couch when she could not move her right arm and leg for several minutes, without associated disturbance of cognition or sensory symptoms. Examination at the emergency room after recovery was normal. Brain imaging was not performed because of the pregnancy. Two subsequent EEGs were normal, and no specific diagnosis was made at the time. She had a history of migraine, and we cannot definitively exclude the possibility that this first episode of unilateral weakness was a migraine aura without headache. However, her typical migraine episodes are headache and are distinct from this presentation. Fifteen years later, while driving, she noted weakness of her arms, legs, and trunk severe enough to warrant stopping the car. Weakness recovered slowly after 15 minutes. A similar episode occurred a few months later. Potassium levels and clinical examination when asymptomatic were normal. Subsequently, the frequency of similar episodes with flaccid paresis of her limbs, affecting predominantly the legs and lasting several minutes, increased and they occurred daily, always after resting. No correlation with food intake or fasting was reported. In addition, she complained of painless cramps in her arms and legs muscles, which could become painful if she tried to stretch her muscles. These lasted up to 10 minutes and also occurred during sleep.

A detailed neurologic examination at the age of 42 years revealed no abnormalities. A relatively thin and long face with a small lower jaw was noted, but she was not overtly dysmorphic. On follow-up examination, she presented with flaccid weakness of her right hand lasting for 2 days. Motor and sensory nerve conduction studies, repetitive motor nerve stimulation, and EMG in the limbs and paravertebral muscles performed after recovery from the acute episode were unremarkable. The McManis test was negative. Laboratory testing revealed no thyroid dysfunction; electrolytes and creatine kinase were normal. Muscle biopsy of the tibialis anterior muscle was unremarkable. Genetic testing revealed *RYR1* gene variants Arg1507Gln and Gly2446Ser in trans. Arg1507Gln is a missense variant with a minor allele frequency of 0.00001 in ExAC that has been previously found in other myopathic phenotypes. Gly2446Ser, although not previously reported, localizes to a recognized MH-associated mutational hot spot ([www.emhg.org](http://www.emhg.org)). PP gene testing was negative.

### Case 3

This 49-year-old man reported experiencing episodes of minor limb weakness after strenuous exercise that lasted a few hours from the age of 14 years. He had his first full attack of muscle paralysis at the age of 29 years. He awoke in the morning after a day of strenuous exercise to find that he was unable to walk. From this age on, he experienced multiple similar episodes. All muscles from the neck down could be weak, although the legs were predominantly and most severely affected. Symptoms usually lasted for several hours

but could persist for as long as 48 hours. In the recovery phase, he reported that the limbs could be painful as strength returned. Symptoms could be provoked by exercise, intense heat, or a carbohydrate-rich meal late at night.

Medical history was notable for 2 complicated general anesthetics in childhood, although the exact circumstances were unclear and medical records unavailable. His parents were advised after these that he may be at risk of MH and to inform clinicians before any future procedures requiring general anesthesia.

Examination was unremarkable, as was creatine kinase and MRI of the lower limbs. EMG and nerve conduction study were normal, but the McManis test for PP was positive with an exercise-induced reduction of CMAP of 68%. A biopsy of the tibialis anterior muscle demonstrated variation in fiber size, an increase in internal nuclei, and 1 ring fiber (figure e-1, <http://links.lww.com/WNL/A96>). A type I fiber predominance was interpreted as normal for the muscle biopsied. Genetic testing for PP genes was negative, but an *RYR1* variant Arg1043His was identified. Another amino acid substitution at the same site, Arg1043Cys, previously associated with the MH trait but not functionally tested yet ([www.emhg.org](http://www.emhg.org)), has been described.

## Discussion

We have previously reported a single patient with recessive *RYR1*-related MmD with additional episodic muscle weakness/paralysis.<sup>11</sup> Here, we have described 3 additional cases (summarized in the table). One was compound heterozygous for *RYR1* nonsense and missense mutations and presented with later development of episodic symptoms in the context of a congenital myopathy, similar to our original patient reported by Zhou et al.<sup>17</sup> who showed a comparable genotype and phenotype. The other 2 patients had an episodic phenotype without myopathy and were either heterozygous for or compound heterozygous for *RYR1* missense mutations (putatively) implicated in the MH trait, suggesting tentative genotype-phenotype correlations. In all cases (including our original case), the onset of episodic muscle paralysis was from early adulthood onward, which is later than the onset typically seen in primary PP.<sup>17</sup> Although there were variable triggers, including exertion or rest after exertion, which is a typical trigger for PP, there was no consistent relationship to food or temperature across all the cases. Myalgia or cramp was a universal feature, indicating overlap with the spectrum of *RYR1*-related exertional myalgia/rhabdomyolysis, and this, together with the later age at onset, may be a phenotypic clue for future cases.

The McManis test was positive in 2 of 3 of our new cases. We retrospectively reviewed the source data for our previously reported case<sup>11</sup> and found that the drop in CMAP amplitude when calculated from the postexercise increment was also >40%, indicating a positive test.

**Table** Summary of clinicopathologic features of cases with *RYR1* gene variants and episodic weakness/paralysis

	Symptoms	Examination	CK, IU/L	EMG	McManis test	Muscle biopsy	<i>RYR1</i> variants
<b>Previously reported case<sup>11</sup></b>	Hypotonia, respiratory and feeding impairment at birth; delayed motor milestones; episodic limb weakness lasting days from age 18 y; cold- and exercise-induced myalgia/cramp from age 26 y	High arched palate; facial weakness; ophthalmoplegia; severe trunk and neck weakness; moderate limb weakness; hyperlordosis	Normal range	Myopathic	Positive	Variation in fiber size; increased internal nuclei; core-like structures	Asp708Asn; Arg2241X; Arg2939Lys
<b>Case 1</b>	Delayed walking; early onset proximal myopathy; episodic myalgia followed by limb and neck weakness lasting hours to days from age 34 y; fluctuant bulbar weakness	Long thin face; high arched palate; proximal myopathy; facial weakness and ptosis; ophthalmoplegia; hyperlordosis	101	Myopathic; increased jitter and blocking on SFEMG	Positive	Variation in fiber size; increased internal nuclei; areas of central pallor suggestive of core-like structures	Gln70X; Arg109Trp; Met485Val
<b>Case 2</b>	Early history and development normal; episodic limb weakness from age 23 y; cramps with myalgia	Long thin face	120	Normal	Negative	Normal	Arg1507Gln; Gly2446Ser
<b>Case 3</b>	Early history and development normal; minor episodic weakness from age 14 y; episodic limb paralysis and myalgia from age 29 y	Normal	92	Normal	Positive	Type 1 predominance; 1 ragged red fiber (+COX negative); increased internal nuclei; 1 ring fiber	Arg1043His

Abbreviations: CK = creatine kinase; SFEMG = single-fiber EMG.

A positive McManis test, to the best of our knowledge, has previously been described only in primary or secondary PP.<sup>18</sup> Given the direct interaction between the Cav1.1 channel implicated in the majority of cases of hypokalemic PP<sup>19</sup> and *RYR1*, it seems plausible that dysfunction of each may produce phenotypic mimics. This is supported by the observation of a profound disturbance of the normal Cav1.1/*RYR1* interaction in 1 previous patient with *RYR1*-related PP and other patients with recessive *RYR1*-related myopathies.<sup>11,20</sup> Furthermore, the recently described *CACNAIS*-associated early-onset myopathy shares features of predominantly axial weakness and ophthalmoplegia with *RYR1*-related myopathies<sup>12</sup> and may also feature similar reductions of the Cav1.1 protein. However, the exact mechanism by which the *RYR1* variants would cause an episodically unexcitable sarcolemma as demonstrated by a positive McManis test is unclear. On the basis of the recent observation of altered BK channel activity in *RYR1*-mutated smooth muscle cells,<sup>10</sup> one possibility is via an impairment of BK channel-mediated membrane repolarization due to altered intracellular calcium homeostasis, but this requires further study.<sup>10,21,22</sup>

Other atypical PP phenotypes associated with motor neuropathy due to mitochondrial gene mutations have been described,<sup>23</sup> demonstrating that the clinical symptoms may be seen in non-ion channel genetic disorders. Our data demonstrate that late-onset episodic muscle weakness or paralysis with prominent myalgia and cramps may be a more common *RYR1*-associated phenotype than the single case we previously

reported and may not be limited to a specific genotype. We suggest that cases compound heterozygous for *RYR1* nonsense and missense mutations are more likely to have a concomitant early-onset myopathy. The McManis test may be a useful diagnostic tool in these cases, although a negative test does not exclude the possibility of *RYR1* involvement.

We propose that clinicopathologic features suggestive of *RYR1* disorders should be sought in genetically undefined PP cases and *RYR1* gene testing considered in those in whom mutations in *SCN4A*, *CACNAIS*, and *KCNJ2* have already been excluded.

### Author contributions

E. Matthews: drafting the manuscript for content, analysis or interpretation of data, study concept. C. Neuwirth and F. Jaffer: revising the manuscript for content, analysis or interpretation of data, study concept. R. Scalco, D. Fialho, M. Parton, R. Sud, R. Spiegel, R. Mein, D. Raja Rayan, K. Suetterlin, H. Houlden, A. Schaefer, E. Healy, J. Palace, R. Quinlivan, S. Treves, and J.L. Holton: revising the manuscript for content, analysis or interpretation of data. H. Jungbluth: drafting the manuscript for content, analysis or interpretation of data, study concept. M.G. Hanna: revising the manuscript for content, analysis or interpretation of data.

### Acknowledgment

The authors are grateful to Mark Walker at Southampton for the retrieval of muscle histology slides.

## Study funding

Part of this work was undertaken at University College London Hospitals/University College London, which received a proportion of funding from the Department of Health's National Institute for Health Research Biomedical Research Centres funding scheme.

## Disclosure

E. Matthews holds a post-doctoral fellowship from the National Institute for Health Research Rare Disease Scheme. C. Neuwirth, F. Jaffer, and R. Scalco report no disclosures relevant to the manuscript. D. Fialho is supported by the National Highly Specialised Service Department of Health UK. M. Parton, R. Sud, R. Spiegel, R. Mein, and D. Rayan report no disclosures relevant to the manuscript. K. Suetterlin has an MRC Clinical Research Training Fellowship and received funding from the European Community's Seventh Framework Programme (FP7/2007-2013) under grant agreement 2012-305121 "Integrated European-omics Research Project for Diagnosis and Therapy in Rare Neuromuscular and Neurodegenerative Diseases (NEUROMICS)". H. Houlden has received funding from the European Community's Seventh Framework Programme (FP7/2007-2013) under grant agreement 2012-305121 "Integrated European-omics Research Project for Diagnosis and Therapy in Rare Neuromuscular and Neurodegenerative Diseases (NEUROMICS)". A. Schaefer and E. Healy report no disclosures relevant to the manuscript. J. Palace is partly funded by Highly Specialised Services to run national congenital myasthenia and neuromyelitis optica services and has received support for scientific meetings and honorariums for advisory work from Merck Serono, Biogen Idec, Novartis, Teva, Chugai Pharma, and Bayer Schering, as well as unrestricted grants from Merck Serono, Novartis, Biogen Idec, Teva, and Bayer Schering. Her hospital trust receives funds for her role as clinical lead for the Department of Health Risk Sharing Scheme. She has received grants from the MS society and Guthy-Jackson Foundation. R. Quinlivan is supported by the National Highly Specialised Service Department of Health UK and National Institute for Health Research and has grant funding from Muscular Dystrophy UK and Association for Glycogen Storage Disease. She has received honoraria for teaching and consultancy from PTC Therapeutics, Sanofi-Genzyme, and Ultragenyx. S. Treves reports no disclosures relevant to the manuscript. J. Holton is supported by the Reta Lila Weston Institute for Neurologic Studies. H. Jungbluth reports no disclosures relevant to the manuscript. M. Hanna is

supported by a Medical Research Council Centre grant, the National Centre for Research Resources, and the National Highly Specialised Service Department of Health UK. Go to [Neurology.org/N](http://Neurology.org/N) for full disclosures.

Received June 9, 2017. Accepted in final form October 24, 2017.

## References

1. Amburgey K, McNamara N, Bennett LR, McCormick ME, Acsadi G, Dowling JJ. Prevalence of congenital myopathies in a representative pediatric United States population. *Ann Neurol* 2011;70:662–665.
2. Maggi L, Scoto M, Cirak S, et al. Congenital myopathies: clinical features and frequency of individual subtypes diagnosed over a 5-year period in the United Kingdom. *Neuromuscul Disord* 2013;23:195–205.
3. Zhang Y, Chen HS, Khanna VK, et al. A mutation in the human ryanodine receptor gene associated with central core disease. *Nat Genet* 1993;5:46–50.
4. Jungbluth H, Zhou H, Hartley L, et al. Minicore myopathy with ophthalmoplegia caused by mutations in the ryanodine receptor type 1 gene. *Neurology* 2005;65:1930–1935.
5. Wilmshurst JM, Lillis S, Zhou H, et al. RYR1 mutations are a common cause of congenital myopathies with central nuclei. *Ann Neurol* 2010;68:717–726.
6. Clarke NF, Waddell LB, Cooper ST, et al. Recessive mutations in RYR1 are a common cause of congenital fiber type disproportion. *Hum Mutat* 2010;31:E1544–E1550.
7. Dlamini N, Voermans NC, Lillis S, et al. Mutations in RYR1 are a common cause of exertional myalgia and rhabdomyolysis. *Neuromuscul Disord* 2013;23:540–548.
8. Jungbluth H, Lillis S, Zhou H, et al. Late-onset axial myopathy with cores due to a novel heterozygous dominant mutation in the skeletal muscle ryanodine receptor (RYR1) gene. *Neuromuscul Disord* 2009;19:344–347.
9. Loseth S, Voermans NC, Torbergesen T, et al. A novel late-onset axial myopathy associated with mutations in the skeletal muscle ryanodine receptor (RYR1) gene. *J Neurol* 2013;260:1504–1510.
10. Lopez RJ, Byrne S, Vukcevic M, et al. An RYR1 mutation associated with malignant hyperthermia is also associated with bleeding abnormalities. *Sci Signal* 2016;9:ra68.
11. Zhou H, Lillis S, Loy RE, et al. Multi-minicore disease and atypical periodic paralysis associated with novel mutations in the skeletal muscle ryanodine receptor (RYR1) gene. *Neuromuscul Disord* 2010;20:166–173.
12. Schartner V, Romero NB, Donkersvoort S, et al. Dihydropyridine receptor (DHPR, CACNA1S) congenital myopathy. *Acta Neuropathol* 2017;133:517–533.
13. Illingworth MA, Main M, Pitt M, et al. RYR1-related congenital myopathy with fatigable weakness, responding to pyridostigmine. *Neuromuscul Disord* 2014;24:707–712.
14. Jungbluth H, Davis MR, Muller C, et al. Magnetic resonance imaging of muscle in congenital myopathies associated with RYR1 mutations. *Neuromuscul Disord* 2004;14:785–790.
15. Klein A, Jungbluth H, Clement E, et al. Muscle magnetic resonance imaging in congenital myopathies due to ryanodine receptor type 1 gene mutations. *Arch Neurol* 2011;68:1171–1179.
16. Zhou H, Jungbluth H, Sewry CA, et al. Molecular mechanisms and phenotypic variation in RYR1-related congenital myopathies. *Brain* 2007;130:2024–2036.
17. Venance SL, Cannon SC, Fialho D, et al. The primary periodic paralyses: diagnosis, pathogenesis and treatment. *Brain* 2006;129:8–17.
18. McManis PG, Lambert EH, Daube JR. The exercise test in periodic paralysis. *Muscle Nerve* 1986;9:704–710.
19. Matthews E, Labrum R, Sweeney MG, et al. Voltage sensor charge loss accounts for most cases of hypokalemic periodic paralysis. *Neurology* 2009;72:1544–1547.
20. Zhou H, Rokach O, Feng L, et al. RyR1 deficiency in congenital myopathies disrupts excitation-contraction coupling. *Hum Mutat* 2013;34:986–996.
21. Tricarico D, Mele A, Conte CD. Carbonic anhydrase inhibitors ameliorate the symptoms of hypokalaemic periodic paralysis in rats by opening the muscular Ca<sup>2+</sup>-activated-K<sup>+</sup> channels. *Neuromuscul Disord* 2006;16:39–45.
22. Kim JB, Kim SJ, Kang SY, Yi JW, Kim SM. The large-conductance calcium-activated potassium channel holds the key to the conundrum of familial hypokalemic periodic paralysis. *Korean J Pediatr* 2014;57:445–450.
23. Aure K, Dubourg O, Jardel C, et al. Episodic weakness due to mitochondrial DNA MT-ATP6/8 mutations. *Neurology* 2013;81:1810–1818.

# Neurology®

## Atypical periodic paralysis and myalgia: A novel RYR1 phenotype

Emma Matthews, Christoph Neuwirth, Fatima Jaffer, et al.

*Neurology* published online January 3, 2018

DOI 10.1212/WNL.0000000000004894

**This information is current as of January 3, 2018**

<b>Updated Information &amp; Services</b>	including high resolution figures, can be found at: <a href="http://n.neurology.org/content/early/2018/01/03/WNL.0000000000004894.full">http://n.neurology.org/content/early/2018/01/03/WNL.0000000000004894.full</a>
<b>Subspecialty Collections</b>	This article, along with others on similar topics, appears in the following collection(s): <b>All Clinical Neurology</b> <a href="http://n.neurology.org/cgi/collection/all_clinical_neurology">http://n.neurology.org/cgi/collection/all_clinical_neurology</a> <b>EMG</b> <a href="http://n.neurology.org/cgi/collection/emg">http://n.neurology.org/cgi/collection/emg</a> <b>Ion channel gene defects</b> <a href="http://n.neurology.org/cgi/collection/ion_channel_gene_defects">http://n.neurology.org/cgi/collection/ion_channel_gene_defects</a> <b>Muscle disease</b> <a href="http://n.neurology.org/cgi/collection/muscle_disease">http://n.neurology.org/cgi/collection/muscle_disease</a>
<b>Permissions &amp; Licensing</b>	Information about reproducing this article in parts (figures, tables) or in its entirety can be found online at: <a href="http://www.neurology.org/about/about_the_journal#permissions">http://www.neurology.org/about/about_the_journal#permissions</a>
<b>Reprints</b>	Information about ordering reprints can be found online: <a href="http://n.neurology.org/subscribers/advertise">http://n.neurology.org/subscribers/advertise</a>

*Neurology*® is the official journal of the American Academy of Neurology. Published continuously since 1951, it is now a weekly with 48 issues per year. Copyright © 2018 The Author(s). Published by Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology. All rights reserved. Print ISSN: 0028-3878. Online ISSN: 1526-632X.

