Neurofilament light protein in blood predicts regional atrophy in Huntington disease

Eileanoir B. Johnson, MSc, Lauren M. Byrne, MSc, Sarah Gregory, PhD, Filipe B. Rodrigues, MD, Kaj Blennow, MD, PhD, Alexandra Durr, MD, PhD, Blair R. Leavitt, MD, Raymund A. Roos, MD, Henrik Zetterberg, MD, PhD, Sarah J. Tabrizi, MBChB, PhD, Rachael I. Scahill, PhD, and Edward J. Wild, MBChB, PhD, for the TRACK-HD Study Group

Neurology® 2018;90:e717-e723. doi:10.1212/WNL.0000000000005005

Abstract

Objective
Neurofilament light (NfL) protein in blood plasma has been proposed as a prognostic biomarker of neurodegeneration in a number of conditions, including Huntington disease (HD). This study investigates the regional distribution of NfL-associated neural pathology in HD gene expansion carriers.

Methods
We examined associations between NfL measured in plasma and regionally specific atrophy in cross-sectional (n = 198) and longitudinal (n = 177) data in HD gene expansion carriers from the international multisite TRACK-HD study. Using voxel-based morphometry, we measured associations between baseline NfL levels and both baseline gray matter and white matter volume; and longitudinal change in gray matter and white matter over the subsequent 3 years in HD gene expansion carriers.

Results
After controlling for demographics, associations between increased NfL levels and reduced brain volume were seen in cortical and subcortical gray matter and within the white matter. After also controlling for known predictors of disease progression (age and CAG repeat length), associations were limited to the caudate and putamen. Longitudinally, NfL predicted subsequent occipital gray matter atrophy and widespread white matter reduction, both before and after correction for other predictors of disease progression.

Conclusions
These findings highlight the value of NfL as a dynamic marker of brain atrophy and, more generally, provide further evidence of the strong association between plasma NfL level, a candidate blood biomarker, and pathologic neuronal change.
The identification of sensitive biomarkers of disease progression could help in the development of successful treatments for neurodegenerative conditions. Blood biomarkers are particularly appealing as they provide a quick, noninvasive, objective, and reproducible way of quantifying markers of disease progression. Neurofilament light (NfL) has been recognized as a CSF biomarker of neuronal damage for a number of years, but recent advances in ultrasensitive immunoassays have enabled quantification of NfL from blood samples. Recently, NfL was proposed as a candidate prognostic blood biomarker of Huntington disease (HD) onset and progression. Baseline NfL was retrospectively quantified in plasma using the well-characterized TRACK-HD cohort. NfL was increased in HD gene expansion carriers when compared to controls, with NfL levels remaining elevated long after disease onset, were calculated for this cohort. Participants unclassified as having premanifest HD (preHD) and 123 participants with HD were recruited as part of the multisite longitudinal TRACK-HD study, which included 120 participants with premanifest HD (preHD) and 123 participants with manifest HD. Participants were recruited in 2008 from hospital clinics at 4 sites based in Leiden, London, Paris, and Vancouver and were classified as having premanifest or manifest HD at baseline depending on their score on the Unified Huntington’s Disease Rating Scale (UHDRS) Total Motor Score. A score of ≤5 meant that a participant was categorized as preHD, and participants with a Total Motor Score >5 combined with a UHDRS Total Functional Capacity score >7 were classed as early manifest HD. Disease Burden Score, an approximate marker of disease load, and 5-year probability of onset, an approximate prediction of disease onset, were calculated for this cohort. Participants underwent a series of assessments and returned yearly until 2011. Sample size was calculated before the TRACK-HD baseline visit and was aimed at detecting significant longitudinal change over 2 years in different variables. Full study and recruitment information has been documented previously.

**Standard protocol approvals, registrations, and patient consents**

Local ethical approval was given for the study and all participants gave their written informed consent according to the Declaration of Helsinki.

**Imaging acquisition and processing**

Three-tesla MRI scans were collected at all 4 sites using a standardized T1-weighted acquisition developed for this study. Scans underwent rigorous quality control, with meta-data checks and visual quality-control steps performed to ensure that acquisition parameters were correct and to exclude scans with motion and other artifacts. All scans that passed quality control were then processed using SPM 12 (http://www.fil.ion.ucl.ac.uk/spm/) with MATLAB version R2012B (https://in.mathworks.com). Baseline volumes were separated into different tissue classes (gray matter [GM], white matter [WM], and CSF) using the Segment procedure. A group template was created using Diffeomorphic Anatomical Registration Through Exponentiated Lie Algebra (DARTEL), and the WM and GM tissue classes were then warped to the DARTEL template. These images were modulated and smoothed to account for any volume changes that occurred during normalization. A 4-mm kernel at full-width half maximum was used for smoothing. All images were visually examined to check segmentation and normalization performance.

**Longitudinal VBM was performed as described previously.** Within-subject longitudinal change was first measured using a nonlinear fluid registration technique within inhouse MIDAS software. Voxel-compression maps were generated for each participant from baseline to 3-year follow-up. These voxel-compression maps were then spatially mapped onto the DARTEL template and then convolved with the participant-specific baseline GM and WM maps to generate voxel-level, within-subject change for each tissue type.

Total intracranial volume was measured via the MIDAS software using a semiautomated procedure as previously described. All image analyses were performed blinded to participant group to avoid potential sources of bias.

**Methods**

**Participants**

Participants were recruited as part of the multisite longitudinal TRACK-HD study, which included 120 participants with premanifest HD (preHD) and 123 participants with manifest HD. Participants were recruited in 2008 from hospital clinics at 4 sites based in Leiden, London, Paris, and Vancouver and were classified as having premanifest or manifest HD at baseline depending on their score on the Unified Huntington’s Disease Rating Scale (UHDRS) Total Motor Score. A score of ≤5 meant that a participant was categorized as preHD, and participants with a Total Motor Score >5 combined with a UHDRS Total Functional Capacity score >7 were classed as early manifest HD. Disease Burden Score, an approximate marker of disease load, and 5-year probability of onset, an approximate prediction of disease onset, were calculated for this cohort. Participants underwent a series of assessments and returned yearly until 2011. Sample size was calculated before the TRACK-HD baseline visit and was aimed at detecting significant longitudinal change over 2 years in different variables. Full study and recruitment information has been documented previously.

**Glossary**

DARTEL = Diffeomorphic Anatomical Registration Through Exponentiated Lie Algebra; GM = gray matter; HD = Huntington disease; NfL = neurofilament light; preHD = premanifest Huntington disease; UHDRS = Unified Huntington’s Disease Rating Scale; VBM = voxel-based morphometry; WM = white matter.
NfL quantification
NfL was quantified in plasma samples using an ultrasensitive single-molecule array method as previously described. All samples were analyzed in one round of experiments using one batch of reagents. All NfL values were within the linear ranges of the assays, and intra-assay coefficients of variation were below 10%. NfL quantification was performed blinded to group to avoid potential sources of bias.

Statistical analysis
Associations between NfL and (1) cross-sectional GM or WM volumes, and (2) longitudinal GM and WM change were examined using linear regression models within SPM 12. Age, sex, study site, and total intracranial volume were initially controlled for in the model, to permit identification of regions where atrophy was associated with plasma NfL. To identify regions where NfL independently predicted atrophy, after adjustment for known predictors of HD progression, the analysis was then repeated with CAG repeat length and age–CAG interaction as terms. These terms are known predictors of HD disease onset and stage, and thus by controlling for these factors, we can better understand the independent influence of NfL on GM and WM. Explicit binary masks for GM and WM were used in the analysis, as described previously. All results were one-tailed and corrected for multiple comparisons at \( p = 0.05 \) using family-wise error.

Results
Of the 243 gene expansion carriers recruited in TRACK-HD, 201 had plasma samples collected at baseline (2008) from which NfL could be measured, and 198 also had structural imaging that passed quality control at baseline. Of these participants, 176 had both plasma samples and structural MRI scans at the 3-year follow-up (2011), with an additional participant who had a structural MRI scan but no plasma. Although retention was high for the study, the most common reason for drop-out at the 2011 time point was worsening of symptoms. Detailed information on retention was discussed previously. Demographic information is provided in the table.

### Table demographic information for all participants at baseline

<table>
<thead>
<tr>
<th></th>
<th>Combined</th>
<th>preHD</th>
<th>HD</th>
</tr>
</thead>
<tbody>
<tr>
<td>No.</td>
<td>198</td>
<td>104</td>
<td>94</td>
</tr>
<tr>
<td>Age, y</td>
<td>44.59 (9.96); 18.63–64.13</td>
<td>40.98 (8.67); 18.63–64.13</td>
<td>48.59 (9.81); 22.76–64.11</td>
</tr>
<tr>
<td>Sex, M/F</td>
<td>107/91</td>
<td>56/48</td>
<td>51/43</td>
</tr>
<tr>
<td>CAG</td>
<td>43.32 (2.75); 39.00–59.00</td>
<td>43.04 (2.32); 39.00–50.00</td>
<td>43.64 (3.13); 39.00–59.00</td>
</tr>
<tr>
<td>TMS</td>
<td>12.12 (12.57); 0.00–47.00</td>
<td>2.53 (1.68); 0.00–8.00</td>
<td>22.72 (10.72); 5.00–47.00</td>
</tr>
<tr>
<td>TFC</td>
<td>12.03 (1.63); 7.00–13.00</td>
<td>12.84 (0.59); 9.00–13.00</td>
<td>11.13 (1.91); 7.00–13.00</td>
</tr>
<tr>
<td>DBS</td>
<td>330.30 (74.87); 156.19–566.46</td>
<td>292.63 (47.48); 171.00–408.90</td>
<td>371.98 (77.68); 156.19–566.46</td>
</tr>
<tr>
<td>Probability 5-y onset*</td>
<td>0.34 (0.19); 0.03–0.74</td>
<td>0.22 (0.12); 0.03–0.53</td>
<td>0.46 (0.16); 0.03–0.74</td>
</tr>
<tr>
<td>GM volume, % of TIV</td>
<td>45.42 (4.27); 32.41–56.07</td>
<td>47.57 (3.07); 37.42–56.07</td>
<td>43.04 (4.16); 32.41–51.63</td>
</tr>
<tr>
<td>WM volume, % of TIV</td>
<td>28.74 (2.12); 23.63–34.30</td>
<td>29.57 (1.91); 25.02–34.30</td>
<td>27.82 (1.95); 23.63–33.78</td>
</tr>
</tbody>
</table>

Abbreviations: CAG = length of genetic repeat; DBS = Disease Burden Score; GM = gray matter; HD = Huntington disease; preHD = premanifest HD; TFC = Total Functional Capacity; TIV = total intracranial volume; TMS = Total Motor Score; WM = white matter.
Values are mean (SD); range.
* Conditional probability of onset within 5 years.
significantly in the corpus callosum (figure 3B). Figure 4A shows that after controlling for CAG repeat length and the interaction between age and CAG repeat length, NfL primarily predicted subsequent GM reduction within the inferior occipital cortex and the lateral occipital cortex. Significant associations were also seen in small regions within the inferior frontal gyrus and superior temporal gyrus. Finally, higher baseline NfL again showed widespread associations with longitudinal WM volume reductions, even after adjusting for CAG and the age × CAG interaction term (figure 4B).

**Discussion**

This study presents unbiased whole-brain results showing voxel-wise region-specific associations between NfL levels measured in plasma and both cross-sectional and longitudinal MRI GM and WM volume in a pathologic group previously shown to have raised NfL levels. One previous study in HD found associations between the level of NfL in plasma and MRI measures of cross-sectional brain volume and brain atrophy over the subsequent 3 years in a number of predefined
regions of interest using global values. Here, we build on this previous work, using VBM to reveal the location and extent of volumetric change in brain regions that are associated with NfL levels in HD.

As expected, higher NfL in plasma was associated with lower volume in regions known to be affected in HD. After adjustment for CAG repeat length and its interaction with age—known predictors of HD progression—NfL remained independently associated with reduced GM volume in the caudate and putamen bilaterally, but there was no association with cortical or WM volume at baseline. The caudate and putamen are regions that undergo the earliest and most extensive atrophy in HD due to the predominance of medium spiny neurons that are particularly vulnerable to the effects of mHTT. However, it appears that baseline NfL level does not specifically relate to more widespread neuronal damage at a given time point, beyond the overall general effect of age and CAG repeat length on neurodegeneration. That the caudate and putamen were associated with NfL concentration after age and CAG adjustment suggests that NfL is capable of identifying individuals who, over the course of their life up to the time of sampling, have experienced relatively larger or smaller volume loss in these disease-linked brain regions.

It is of interest that there was a much stronger relationship between baseline NfL levels and subsequent regional volume loss over time, with these relationships remaining highly significant after adjusting for CAG repeat length and its interaction with age. Higher baseline levels of NfL significantly predicted increased atrophy in cortical but not subcortical regions of the GM over the following 3 years, which is consistent with the previous study reporting stronger association between baseline NfL and total GM than caudate atrophy. Our results extend this finding by providing further evidence that, after controlling for overall age and CAG effects, higher NfL is predictive of HD gene expansion carriers who will subsequently undergo greater GM atrophy extending beyond the striatum. This study demonstrates that the most significant associations between NfL and subsequent neuronal change are localized to the occipital lobe, one of the earliest cortical regions thought to undergo atrophy in HD. While previous studies have found that the occipital lobe appears to be an early region of neural change, neuropathology studies have found that in more advanced cases of HD, cortical atrophy is globally distributed. The significant association between higher NfL and subsequent atrophy suggests that NfL is sensitive to some of the earliest cortical atrophy in HD.

In addition to cortical GM atrophy, there was a strong relationship between higher baseline NfL levels and widespread atrophy within the WM. WM has been shown to undergo change in both preHD and manifest HD, and is strongly
related to disease progression. Our findings suggest that, similarly to GM, NfL is predictive of subsequent WM atrophy. These associations indicate that it in addition to its value as an accessible indicator of global neurodegeneration, NfL may have particular value as a marker of axonal degeneration—and therefore potentially reversible neuropathology in regions vulnerable to damage in HD.

Whole-brain analyses using VBM have only been used to examine the relationship between NfL—measured in CSF, not plasma—and neuronal change in a neurologic illness (frontotemporal dementia) in one previous study. However, the current findings are the first to show significant associations between NfL in plasma across the whole brain in both cross-sectional and longitudinal data. Even though plasma NfL has been shown to increase longitudinally in HD, and the associations of such longitudinal change may be of interest, our results suggest that a single baseline measurement of NfL has the ability to identify gene expansion carriers who have previously undergone disproportionate atrophy in the basal ganglia, and those who are likely to go on to develop similarly disproportionate atrophy in cortical and WM regions vulnerable to HD pathology. This corroborates previous findings and supports the hypothesis that NfL is a dynamic, rapidly-assessed marker of ongoing neuronal damage. Previous reports from the TRACK-HD study demonstrated that patients with the highest atrophy rates were those who subsequently showed the greatest clinical decline. Here, we demonstrate that the region-specific predictive power of NfL extends beyond predefined large volumes of interest. This provides further evidence for the utility of NfL as a potential efficacy and disease progression biomarker in therapeutic intervention trials.

We showed a highly significant and widespread association between plasma NfL and change in WM volume using VBM. In the future, use of microstructural measures of WM degeneration derived from diffusion imaging may allow a detailed characterization of the relationship with NfL, providing further mechanistic insights into the breakdown of brain connectivity, which we know is a key feature of HD. Because the progression of neuronal atrophy beyond the caudate in preHD and manifest HD is fairly slow, analysis using longer time intervals and a wider range of disease stages would be useful to further establish the relationship between NfL and neurodegeneration. Application of these techniques in other disease cohorts will help us to understand the wider role of NfL in the neurodegenerative process. Furthermore, this study is limited because the analysis was performed on a group of HD gene expansion carriers across a range of premanifest and manifest stages. This provides more power to detect effects; however, further analyses within subgroups could provide detailed information on different stages of the disease. Finally, because of individual differences among participants,
our findings cannot be applied meaningfully to individual HD mutation carriers or for clinical decision-making.

We provide further evidence supporting the use of NfL as a prognostic marker of progression of neuronal damage in both HD and other neurodegenerative diseases. NfL appears to be a significant indicator of subsequent widespread brain changes extending beyond the striatum, particularly within the WM. The ability to measure NfL from plasma provides an easily accessible biomarker that has close links to the underlying pathology of HD and shows promise as a dynamic marker of ongoing neuronal change.

**Author contributions**

Eileanoir B. Johnson: study design, data processing, data analysis and interpretation, write-up. Lauren M. Byrne: data processing and critical revision of manuscript. Sarah Gregory: study design, data interpretation, critical revision of manuscript. Filipe B. Rodrigues: data processing and critical revision of manuscript. Kaj Blennow: data processing and critical revision of manuscript. Alexandra Durr: site principal investigator for TRACK-HD study and critical revision of manuscript. Blair R. Leavitt: site principal investigator for TRACK-HD study and critical revision of manuscript. Raymund A. Roos: site principal investigator for TRACK-HD study and critical revision of manuscript. Henrik Zetterberg: data processing and critical revision of manuscript. Rachael I. Scahiil: study design, data processing and interpretation, critical revision of manuscript. Edward J. Wild: study concept and design, data interpretation, and critical revision of manuscript.

**Acknowledgment**

The authors thank the TRACK-HD study participants and the CHDI Foundation, a not-for-profit organization dedicated to finding treatments for HD.

**Study funding**

Some of this work was undertaken at University College London Hospital/University College London, which received funding from the Department of Health NIHR Biomedical Research Centres funding scheme. Funding was also provided by CHDI Foundation (Track-HD study and F.R.). The Swedish Research Council (K.B. and H.Z.), the European Research Council (H.Z.), the Torsten Söderberg Foundation (K.B.), the Knut and Alice Wallenberg Foundation (H.Z.), VINNOVA (K.B. and H.Z.), the Wolfson Foundation (H.Z.), Wellcome Trust (E.J., S.G., R.S. and S.T.) grant code: 200181/Z/15/Z, and the Medical Research Council UK (F.R. and E.W.).

**Disclosure**

The authors report no disclosures relevant to the manuscript. Go to Neurology.org/N for full disclosures.

Received September 28, 2017. Accepted in final form November 28, 2017.

**References**

Neurofilament light protein in blood predicts regional atrophy in Huntington disease

Eileanoir B. Johnson, MSc, Lauren M. Byrne, MSc, Sarah Gregory, PhD, Filipe B. Rodrigues, MD, Kaj Blennow, MD, PhD, Alexandra Durr, MD, PhD, Blair R. Leavitt, MD, Raymund A. Roos, MD, Henrik Zetterberg, MD, PhD, Sarah J. Tabrizi, MBChB, PhD, Rachael I. Scahill, PhD, and Edward J. Wild, MBChB, PhD, for the TRACK-HD Study Group

Cite as: Neurology® 2018;90:e717-e723. doi:10.1212/WNL.0000000000005005

Correspondence
Dr. Wild
e.wild@ucl.ac.uk

Study question
How are plasma levels of neurofilament light (NFL) protein related to progressive regional atrophy in people carrying the Huntington disease (HD) gene expansion?

Summary answer
Elevated plasma NFL levels predict occipital gray matter (GM) atrophy and widespread white matter (WM) volume reductions.

What is known and what this paper adds
Plasma NFL levels are a biomarker for HD-related deficits and overall disease progression. This study clarifies the specific regional atrophy patterns that plasma NFL levels can predict.

Participants and setting
This study retrospectively analyzed data for 243 people carrying the HD gene expansion. This included 120 patients with premanifest HD and 123 with manifest HD. All were participants in the TRACK-HD study conducted in London, Leiden, Paris, and Vancouver from 2008 to 2011.

Design, size, and duration
Plasma NFL levels were quantified with an ultrasensitive single-molecule array method. All patients underwent MRI assessments of GM and WM volumes. Cross-sectional and longitudinal voxel-based morphometry were used to analyze regional volume changes over the 3-year study period.

Primary outcomes
The primary outcomes were associations between plasma NFL levels and longitudinal changes in regional GM and WM volumes.

Main results and the role of chance
After adjustments for gene expansion lengths and age × gene expansion length interactions, elevated baseline plasma NFL levels were associated with bilaterally reduced baseline caudate nucleus and putamen volumes. Elevated baseline plasma NFL levels were also associated with widespread longitudinal WM volume reductions and with longitudinal GM volume reductions within the inferior and lateral occipital cortices and small regions of the inferior frontal and superior temporal gyri.

Bias, confounding, and other reasons for caution
The study follow-up length might not have been long enough to fully characterize the relationship between plasma NFL levels and neurodegeneration.

Generalizability to other populations
Interindividual variation prevents these results from being used in clinical decision-making for individual patients. Further studies within the premanifest and manifest HD subgroups are needed to fully understand the clinical importance of plasma NFL levels. These results support the utility of plasma NFL as a biomarker that predicts brain atrophy relevant to neurodegeneration.

Study funding/potential competing interests
This study was funded by the UK, Swedish, and EU governments and by various medical research foundations. The authors report no competing interests. Go to Neurology.org/N for full disclosures.
Neurofilament light protein in blood predicts regional atrophy in Huntington disease
Eileanoir B. Johnson, Lauren M. Byrne, Sarah Gregory, et al.

Neurology 2018;90:e717-e723 Published Online before print January 24, 2018
DOI 10.1212/WNL.0000000000005005

This information is current as of January 24, 2018

Updated Information & Services
including high resolution figures, can be found at:
http://n.neurology.org/content/90/8/e717.full

References
This article cites 23 articles, 3 of which you can access for free at:
http://n.neurology.org/content/90/8/e717.full#ref-list-1

Citations
This article has been cited by 2 HighWire-hosted articles:
http://n.neurology.org/content/90/8/e717.full##otherarticles

Subspecialty Collections
This article, along with others on similar topics, appears in the following collection(s):
Basal ganglia
http://n.neurology.org/cgi/collection/basil_ganglia
Huntington’s disease
http://n.neurology.org/cgi/collection/huntingtons_disease
MRI
http://n.neurology.org/cgi/collection/mri

Permissions & Licensing
Information about reproducing this article in parts (figures, tables) or in its entirety can be found online at:
http://www.neurology.org/about/about_the_journal#permissions

Reprints
Information about ordering reprints can be found online:
http://n.neurology.org/subscribers/advertise

Neurology® is the official journal of the American Academy of Neurology. Published continuously since 1951, it is now a weekly with 48 issues per year. Copyright © 2018 The Author(s). Published by Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology. All rights reserved. Print ISSN: 0028-3878. Online ISSN: 1526-632X.